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Systemic Uptake and Clearance of Chloroform by Hairless Rats Following Dermal Exposure: I. Brief Exposure to Aqueous Solutions

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The animals used in this study were handled in accordance with the principles stated in the *Guide for the Care and Use of Laboratory Animals*, Institute of Laboratory Animal Resources, National Research Council, National Academy Press, 1996, and the Animal Welfare Act of 1966, as amended.

This report has been reviewed by the Office of Public Affairs (PA) and is releasable to the National Technical Information Service (NTIS). At NTIS, it will be available to the general public, including foreign nations.

This technical report has been reviewed and is approved for publication.

FOR THE COMMANDER

TERRY A. CHILDRESS, Lt Col, USAF, BSC
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13. ABSTRACT (Maximum 200 words)

The systemic uptake of chloroform from dilute aqueous solutions into live hairless rats under conditions simulating dermal environmental exposure was studied. Whole blood was sampled during a 30-minute immersion of an animal within water containing a known concentration of chloroform and then for 5.5 hours following its removal from the bath. The amount of chloroform systemically absorbed was determined by comparing the AUCs of the blood concentration versus time plots from dermal exposure to that obtained after IV infusion (for a period of 30 minutes) of an aqueous solution containing a known amount of chloroform (positive control). Although dermal data implied 2-compartment disposition characteristics, IV infusion data fit best to a 3-compartment disposition. Linear pharmacokinetics was observed both by IV administration and percutaneous absorption at the dose levels studied. Chloroform was detected in the rat blood as early as 4 minutes following exposure. Our findings suggest that about 10.2 mg of chloroform was systemically absorbed after dermal exposure of a rat to an aqueous solution of 0.44 mg/ml. This amount is substantially higher than the predictions of risk models put forth by other investigators. Reasonable agreement existed between a past estimate of uptake based on depletion of bath level of chloroform and the actual uptake measured in these experiments. The estimated onset of systemic entry seen here is entirely consistent with our estimate of how long it takes to establish the diffusion gradient across the stratum corneum based on tape stripping.

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1. INTRODUCTION

Recently, there has been a growing concern about the cumulative absorption of chemicals from frequent, intermittent, brief exposures to contaminated water, as might occur daily through bathing[1]. For the most part, such exposures fall short of diffusion lag times for permeation of the skin, which range from several minutes for low molecular weight compounds to several hours for heavier ones [2] and the question has been raised if it is appropriate to use steady-state absorption algorithms to quantify such exposures. The temporal dependencies of the exposures therefore have to be resolved. Chloroform is a representative low molecular weight, lipophilic, very volatile, halogenated compound which is present as a pollutant in our water supplies, and thus a good compound to use to evaluate the unresolved issues. This chemical can be acutely toxic and is also a suspect carcinogen [3]. Therefore, the health risk arising from frequent, brief showering/swimming of human beings in water containing it as a contaminant can not be ignored, especially considering the large surface area of skin which might be exposed.

Although some literature exists on the determination of blood level of chloroform in nonoccupationally exposed humans [4-7] and in animals [8], the means of its intake has generally been other than dermal. Jorgenson and Rushbrook's [8] 90-day subacute study in male rats and female mice, for instance, was aimed at providing toxicologic and range-finding data sufficient for setting dose levels for the chronic-phase testing of chloroform in drinking water. Aggazzotti *et al.* [9] measured plasma chloroform concentrations of swimmers and visitors exposed to chloroform found in the water of three indoor swimming pools in Italy. The blood chloroform concentration depended on both the water and air concentrations, the number of swimmers, the length of swimming time and the intensity of physical activity while in the pool. No effort was made to factor out the dermal contribution. More relevant to the present work, Bogen *et al.*[10] reported dermal uptake of chloroform after exposing hairless guinea pigs to dilute aqueous solutions (10 to 100 µg/L) of chloroform in a closed chamber at 32°C for 70 minutes. Quantification here was indirect and based on measurements of urine and feces concentrations normalized to excretory amounts obtained upon subcutaneous administration of

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chloroform in a corn oil vehicle. Jo *et al.* [11] estimated the chloroform body burden in healthy individuals taking typical showers. Differences in chloroform concentrations in exhaled breath in the course of normal showering and inhalation-only exposures were used to back out the extent of dermal absorption. No direct sampling of blood was attempted. Only a singular investigation seems to have been aimed at obtaining blood concentration-time profiles of chloroform (among other volatile organic chemicals) after its dermal absorption from an aqueous solution, that of Morgan *et al.* [12]. In this work, a small reservoir of the neat chemical or an aqueous solution of it was placed on the back of the animal and kept in place for 24 hours. While all of these studies offer important insights concerning chloroform's uptake, none deals totally effectively with the question of the non-stationary character of uptake associated with very brief exposures.

Earlier [13] we reported on the fate of chloroform taken up into hairless rat skin in the course of exposure to a well-stirred dilute aqueous solution of the chemical. The total dermal uptake was estimated by difference in the bath water concentration of chloroform with and without a rat being present. Measured indirectly, a significant amount of chloroform appeared to reach the viable epidermis and its underlying systemic circulation with exposure times as short as 4 min. The total amount absorbed in a 30-min exposure was estimated to be 29.4 mg. Like studies before it, the indirect method of estimation leaves such conclusions subject to question. To clarify these issues, blood chloroform concentrations arising from short exposure of hairless rats to a dilute aqueous solution have been determined and are reported here.

2. MATERIALS AND METHODS

2.1 Materials and Animals: HPLC grade chloroform (ChromAR® HPLC) was obtained form Mallincrodt, Inc., St. Louis, Missouri. It contained 99% chloroform with 1% ethanol added for stabilization. Hexane used for extraction of chloroform from rat blood was purchased from Fisher Scientific. It was OPTIMA grade and met the rigid analytical requirements for use in electron capture gas chromatography and for residue and pesticide analysis. Since the animals were anesthetized during the experiments, an isothermal pad (Deltaphase™, Braintree Scientific,

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Inc., Braintree, Massachusetts) was placed beneath them to provide warmth while out of the bath. The male hairless rats (CRL:CD) used in the experiments were obtained from Charles River Laboratories, Raleigh, North Carolina. They were 15-20 weeks old, weighing between 410 to 535 g.

2.2 Bath Assembly for Hairless Rats: A tall, 1000 ml, jacketed glass beaker formed the bath in which sedated animals were immersed. This beaker supported a restraining collar (made of stainless steel) used to keep the animal's head above the water and to provide an isolated air environment around the head from which the animal breathed. To accomplish the latter objective, the animal's head was enclosed in a glass chamber formed from an inverted beaker. The rim of this glass chamber fitted into a cylindrical groove, 66 mm in diameter, 4 mm wide and about 2 mm deep, notched into the collar. Air was forced into the chamber through a portal placed over the animal's head. Three symmetrically placed notches across the groove allowed excess to escape at the level of the neck. A photograph of the setup and dimensional details are given our previous paper [13]. A stainless steel rack was placed at the bottom of the beaker to support the rat and allow stirring by a magnetic stirrer placed at the bottom of the vessel. The bath water temperature was held constant at $36 \pm 0.2°C$ (nominally the skin-surface temperature of the rat) using a circulating water bath (Lauda Model M3-B). The entire apparatus was placed inside a fume hood to contain all escaping chloroform.

2.3 Preparation of the Aqueous Solution of Chloroform

2.3.1. Dermal Exposure Solution: Solutions of chloroform in deionized water at 36°C were prepared by adding a calculated volume (0.205 mL) of chloroform to 450 ml of pre-warmed water in a 1000 ml separatory funnel and then vigorously shaking the water for about 2 minutes. Droplets of chloroform at the bottom of the funnel disappeared within this time. This technique ensured complete solution of the test substance and a reproducible initial concentration of 0.44 mg/ml, a concentration 3 orders of magnitude higher than the upper limit of the U. S. EPA toxicity threshold (0.47 µg/ml) [14].

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2.3.2. Positive Control Solution: For the IV experiments, 57 mg (38.41 µl) of chloroform was dissolved in 10 ml of deionized water at 36°C in a 20 ml glass scintillation vial. This solution contained 38 mg (assayed quantity) of chloroform in 10 ml , corresponding to about 50% saturation. Five ml of this solution containing 19 mg of chloroform were intended to be infused into the rat over a 30 min period

2.4. Animal Preparation:

2.4.1. Anesthesia: A combination of Ketamine hydrochloride (87 mg/kg) and Xylazine hydrochloride (13 mg/kg), both given intramuscularly, kept a hairless rat in a sedated state for about 2.5 hours. A second injection was necessary since blood sampling was continued for up to 6 hours. The animals were anesthetized to set the canulae and kept in this state over the entire period of blood sampling.

2.4.2 Jugular Vein Canulation: Because frequent blood sampling took place during the experiments, canulation was done to reduce the stress to the animals. The right jugular vein only was canulated when the rat was dermally exposed to the immersion chamber because only withdrawal of blood samples was required. However, both the right and the left veins had to be canulated for the positive control experiments in order to infuse the control solution of chloroform through the left vein concurrently with blood sampling. The canulae were handprepared from Intramedic PE-50 polyethylene tubing (0.023" ID, 0.034" OD; Beckton Dickinson, Sparks, MD, USA). A small chunk of household adhesive (100% silicone; Dow Corning Corp., Midland, MI) was applied about 1-1/2" from one end of each 7" piece of the tubing and let dry overnight. This bulb of glue allowed us to suture the front portion of the tubing to muscle flanking the incised area. The blunted tip of a 20 G needle was inserted through the other end of the tubing. This served as a port of attachment and detachment of a 1 cc luer tip syringe (used for blood withdrawal). A 1 cc tuberculin syringe was filled with heparinized saline (Heparin conc. - 20 units/ml) and left attached to the hub of this 20 G needle. To set the canula, the ventral skin of the rat above the shoulder was cleaned with an alcohol swab and a vertical, 2 cm long incision was made. After locating the vein, the adjacent connective tissue was cleared

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away and the canula was inserted through the small cut made into the vein by a slanted needle while lifting the vein slightly. The canula was pushed inward towards the heart until blood was seen in the tubing upon pulling back the 1 cc syringe plunger. Some of the heparinized saline was then injected back towards the heart; the syringe was left in place to prevent clotting. Finally, the incision was closed with silk suture.

Exactly the same procedure was followed in canulating the left jugular vein. In order to facilitate the IV infusion of the chloroform solution from an infusion pump into a rat positioned inside the bath, a second piece of polyethylene tubing, about 11" long, was joined to the existing piece through an union made out of the shaft of a 20 G needle.

2.5. IV Administration of the Control Solution: The canulated animal was carefully positioned inside the bath already containing 450 ml of deionized water at 36°C. About 6 ml of the chloroform infusion was withdrawn into a 5 ml Hamilton glass syringe (GASTIGHT® Series 1000) and the luer tip of the syringe was attached to the needle hub from the left canula. The syringe was then inserted into a Harvard Apparatus Syringe Infusion Pump 22 (Harvard Apparatus, Southnatick, MA, USA) and 5 ml solution was infused over 30 min at an infusion rate of 166.7 µl/min. At the end of this period, the infusion line was cut, leaving only about 1" sticking out of the sutured area. The open end of the tubing was sealed with the silicone glue.

2.6. Blood Sampling and Extraction Procedure: In all experiments, one hundred microliters of venous blood were withdrawn through the right canula immediately before putting the animal inside the bath. To prevent chloroform's volatilization, this sample and others to follow were immediately transferred to a 2 ml screw capped autosampler glass vial (New Target DP from Hewlett Packard) containing 1 ml of hexane and vortexed for 1 minute on a Thermolyne Type 16700 Mixer (Barnstead/Thermolyne, Dublique, Iowa). The plastic cap of the vial had a disposable septum with a layer of silicone sandwiched between two linings of Teflon, the latter of which is nonadsorptive to chloroform. The same volume of blood was withdrawn with luer tipped 1 cc Hamilton glass syringes (GASTIGHT® Series 1000) at 4, 8, 20, 30, 45, 60, 90, 120, 180, 240, 300 and 360 min after putting a rat inside the bath. From 0 to 30 min the rat was

actually in the bath. Immediately after the 30-min sample was taken, the animal was taken out of the bath, patted dry with a piece paper towel and laid on the isothermal pad covered with adsorbent paper. The sampling was continued on the time schedule indicated. The entire procedure was performed under a fume hood which provided a laminar flow of room air at 120 - 130 ft/min. The animal's head was positioned so that it was facing the fresh stream of incoming air, eliminating all chance of the animal's inhaling any chloroform.

2.7 Gas Chromatographic Assay : A Hewlett Packard 5890A gas chromatograph equipped with a [63]Ni electron-capture detector, a HP 7673A autosampler, 7673A controller and 3393A integrator was used for the analysis of chloroform. A capillary column (30m x 0.53 mm ID; 1.0 μm film thickness) with DB-17 as the stationary phase obtained from J & W Scientific, Folsom, California, USA was used. Nitrogen was used as the carrier gas with a total flow rate of 120 ml/min (purge vent 20 ml/min, split vent 50 ml/min and column 50 ml/min) and the column head pressure was maintained at 9 psig. The column, the injector and the detector temperatures were set at 65°C, 100°C and 120°C, respectively.

Calibration curves were prepared weekly by plotting the peak areas obtained on injection of 1 μl of the standard solutions against the known concentrations of chloroform in pure hexane. This calibration method was decided upon after extensive work on the assay (to be published separately). One microliter of the clear extract (top hexane layer) of each sample was directly injected into the GC column. The injection was repeated at least two times per sample. The magnetic sample insert of the autosampler was modified to make the injector syringe sample the hexane layer without touching the drop of blood at the bottom of the vial. Between injections the GC injector syringe was rinsed twice with the next sample. Concentrations in the blood samples were determined by dividing the peak areas by the slope of the calibration curve. The standard curves were linear from 10 - 4000 pg/μl and passed through the origin.. Extraction efficiency was determined by equilibrating a series of chloroform-in-hexane solutions with 100 μl of rat blood in 2 ml GC autosampler vials and expressing the chloroform concentration in the hexane

layer found after equilibration as the percent recovered. This proved to be near 100% at concentrations ≥ 100 pg/µl and ranged between 75 and 100% from about 10 to 100 pg/µl.

3. RESULTS AND DISCUSSION

Blood profiles obtained upon infusing 19 mg of chloroform at a steady rate into the jugular vein over 30 min and also after whole body exposure (up to the neck) of a separate set of rats for the same period to a 36°C aqueous solution of chloroform initially containing 0.44 mg/ml of the test pollutant are provided in Figure 1. These curves are composits of the data obtained with 4 animals in cases of both IV infusion and aqueous immersion. The first notable aspect of the profiles is that more than 30 min of steady infusion is obviously necessary to achieve a steady blood level. In other words, neither the IV infusion nor the percutaneous absorption curves reached a peak at the 30-min terminus of the exposure. It is also evident from the inset of Figure 1 that blood levels achieved as the result of dermal absorption lag only a little behind those with IV infusion. The blood levels are very sensitive to input and drop immediately upon cessation of the infusion/exposure processes. In a previous study in which tape stripping was used to measure chloroform in the surface of the skin [13], it was concluded that the gradient of diffusion of chloroform across the stratum corneum is established within 8 min or less, the best estimate by this means being about 4 min. It is evident from the inset plot of Figure 1 that the lag time for systemic breakthrough, that is, the time to systemic entry, is on the order of 4 min by pharmacokinetic analysis. This is a highly satisfying level of agreement between these remarkably different means of estimation.

In order to elucidate the compartmental characteristics of chloroform's absorption and elimination, the natural logarithms of the blood concentrations were plotted against time, as shown in Figure 2. Linear pharmacokinetcs was apparent at the dose levels studied in that neither of the curves shows concavity towards the time axis. At the risk of slightly overprocessing these data, the dermal exposure curve evidences one inflection point indicating a two compartmental disposition following percutaneous input. The IV infusion curve, however, actually evidences two inflection points, implying a three compartmental disposition pattern may

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exist. These essential features were confirmed by applying the method of residuals to both curves [15]. The terminal elimination rate constants (K, min^{-1}) shown in Table I were calculated using the last four data points from each curve. For the dermal exposure, the first-order input rate constant (K_a, min^{-1}) and the first post-absorptive hybrid rate constant (α, min^{-1}), each listed in Table I, were calculated by the method of residuals. In the instance of IV infusion, the first post-absorptive hybrid rate constant (α, min^{-1}) could not be determined because of an insufficiency of data points. A value for this of 0.16 min^{-1} was achieved by overall best-fit analysis, however. PCNONLIN Nonlinear Estimation Program (version 4.2) [16] was used to fit the experimental data to the appropriate models with the blood concentration data being weighted by a factor of 1/concentration. Values for all these rate constants by both methods of estimation are found in Table I along with the estimated time (T_{max}) to reach the maximum blood concentration (C_{max}) values. The following points are notable. The peak time (T_{max}) upon infusion coincided exactly with the moment of termination of the infusion, which was 30 min. This peak appears 1 to 3 minutes later for the data obtained via skin permeation. This slight shift, though not statistically separable, is actually expected because the skin acts as a reservoir for chloroform for a short period. Stripping data indicate this reservoir is depleted in but a few minutes [13]. Also, regardless of which set of data is used and how the calculations are done, the terminal elimination rate constants are all in good agreement, lying between 0.005 and 0.011 min^{-1}.

The areas under the curves (AUCs) were calculated for the blood concentration-time profiles using the trapezoidal rule [15]. To obtain $AUC_{0-\infty}$, the area from the last concentration datum point, C_n, to infinity was set equal to C_n/K and this was added to the area, AUC_{0-C_n}. These parameters too are listed in Table I. The best-fit estimates in the Table are for a two compartment model (first-order absorption with lag time) in the instance of the dermal data but for a three compartment model for the IV infusion data with known zero-order input.

A few investigators have reported pharmacokinetics of chloroform after oral and IV administration in humans [17, 18] and in rodents [19-21]. Fry *et al.* [18] monitored chloroform

concentration in the blood, urine and in expired air as well as the concentration of its metabolite, CO_2, in the expired air of human subjects after oral administration of a capsule containing 500 mg of chloroform in 1 ml olive oil. The blood level data declined in a biphasic fashion similar to what we have found in our dermal exposure experiments. Their reported average (n=4) half-life for the first post-absorptive disposition corresponds to a rate constant of 0.049 min^{-1} which closely matches the value of $\alpha = 0.033$ min^{-1} in hairless rats (n=4) found herein. The average value of the terminal disposition half-life (90 min, n=4), reported in the Fry *et al.*'s study, corresponded to a rate constant of 0.008 min^{-1}, which also closely matched our observed K of 0.009 min^{-1} (Table I) in hairless rats (n=4). Chiou [17] used the data generated by Fry *et al.* to develop equations for predicting the extent of pulmonary and hepatic metabolism of chloroform. Both sources concluded that about 50% of the orally administered chloroform is excreted unchanged in the expired air and the rest metabolized in the liver and excreted as CO_2 through the lungs. Since our dermal exposure experiments with hairless rats resulted in comparable values of the disposition rate constants, it is reasonable to assume that chloroform, absorbed dermally from an aqueous solution of about 0.4 mg/ml, will be metabolized and cleared from the rat body to similar extents as observed with humans after oral administration of 500 mg. Withey *et al.* [19] monitored blood levels of chloroform after intragastric administration of the chemical dissolved in 5 ml of water and equal volume of corn oil. After giving an aqueous solution dose of 75 mg/kg to male Wistar rats (body weight - 400 g), the authors observed saturable elimination kinetics from the log blood concentration-time curves. Although we infused about 42 mg/kg (control experiment) and submerged the hairless rats (body weight - 450 g) in 450 ml water having an initial amount of 200 mg chloroform (dermal exposure experiment), the log blood concentration-time curve clearly demonstrated linear (unsaturable) elimination kinetics (Figure 2). In another study, Withey and Collins [20] reported pharmacokinetics of chloroform in male Wister rats (400 g) after IV bolus administration of 1 ml aqueous solution at five different dose levels (3 - 15 mg/kg). Chloroform reportedly confered three-compartment disposition characteristics within the rat model at all dose levels. These authors found

significantly decreasing values for the disposition rate constants (α, β and γ) with an increase in dose. This would imply dose-dependent disposition kinetics. Michaelis-Menten type metabolism kinetics of chloroform was also observed after inhalation exposure of male Osborne-Mendel rats and male B6C3F1 mice to the radiolabelled chemical [21]. At the highest dose (15 mg/kg) studied by Withey and Collins [20], the model-estimated values for α, β and γ were 0.35 min^{-1}, 0.07 min^{-1} and 0.013 min^{-1}, respectively. These values are systematically higher than the corresponding best-fit values (0.16 min^{-1}, 0.019 min^{-1} and 0.005 min^{-1}) we have obtained (Table I, K represents γ). This is not surprising since, at our IV infusion dose of about 42 mg/kg, one expects about a 2.5-fold decrease in these pharmacokinetic parameters as compared to a 15 mg/kg dose, this expectation being drawn in light of the nonlinear metabolism observed by Withey and Collins [20] and Corley *et. al.* [21].

Since our prime objective in this study was to assess the chloroform body burden arising from its systemic uptake during an exposure to a dilute aqueous solution, we utilized the AUC after IV infusion as the reference standard to quantitate the amount of chloroform absorbed over the 30 min of expsure. We decided to put this calculated systemic uptake value in Table II alongside our previously reported total uptake found *by difference* by direct sampling of the bath water [13] as well as the values predicted by risk assessment models suggested by other investigators [22-24]. Although we determined the systemic uptake only after 30 min exposure in this study by simply using the ratio of AUCs after IV infusion and dermal exposure, the same technique would provide proportional values for shorter dermal exposure times. It is clear that the systemic uptake of chloroform as determined by blood sampling is about three times higher than what we previously found by following the depletion of chloroform from the bath water with and without a rat present. Local tissue binding of chloroform likely plays a role in this quantitative discrepancy. In fact, early investigations confirmed that administration of chloroform to rodents results in a high degree of irreversible binding of nonvolatile metabolites, predominantly in the surface epithelia of the respiratory and upper alimentary tracts and also to a significant extent in the liver and kidney [25-27]. Therefore, it is reasonable to assume that the

AUC from the blood level of unmetabolized chloroform versus time data in our study underestimates, to some extent, the actual amount of chloroform inside the hairless rat's body.

The systemic uptake of chloroform found in this study, however, is badly underestimated by the various models put forth by other investigators [22-24]. This observation clearly indicates that, irrespective of the literature method used to quantitate the body burden of chloroform after dermal exposure to a dilute aqueous solution, the actual risk is far higher. It seems fair to say that these models should be revisited and reworked in light of the present data.

REFERENCES

1. A. L. Bunge, G. L. Flynn and R. H. Guy, "Predictive Model for Dermal Exposure Assessment," in R. G. M. Wang (ed.), *Water Contamination and Health* (Mercel Dekker, New York, 1994).

2. G. L. Flynn, "Physicochemical Determinants of Skin Absorption," in T. R. Gerrity and C. J. Henry (eds.), *Principles of Route-to-Route Extrapolation for Risk Assessment* (Elsevier Science Publishing Company, New York, 1990).

3. S. Budavari (ed.), *The Merck Index*, 11th edition (Merck and Co., Inc., Rahway, New Jersey, 1989).

4. H. Hajimiragha, U. Ewers, R. Jansen-Rosseck and A. Brockhaus, "Human Exposure to Volatile Halogenated Hydrocarbons from the General Environment," *Int. Arch. Occup. Environ. Health* **58**: 141-150 (1986).

5. R. Kroneld, "Chloroform in Tap Water and Human Blood," *Bull. Environ. Contam. Toxicol.* **36**: 477-483 (1986).

6. C. D. Pfaffenberger and A. J. Peoples, "Long-term Variation Study of Blood Plasma Levels of Chloroform and Related Purgeable Compounds," *J. Chromatogr.* **239**: 217-226 (1982).

7. D. L. Ashley, M. A. Bonin, F. L. Cardinali, J. M. McCraw and J. V. Wooten, "Blood Concentrations of Volatile Organic Compounds in a Nonoccupationally Exposed US Population and in Groups with Suspected Exposure," *Clin. Chem.* **40**(7): 1401-1404 (1994).

8. T. A. Jorgenson and C. J. Rushbrook, "Effects of Chloroform in the Drinking Water of Rats and Mice: Ninety-day Subacute Toxicity Study," U.S. Environmental Protection Agency, Report No. EPA-600/1-80-030, July 1980.

9. G. Aggazzotti, G. Fantuzzi, P. L. Tartoni and G. Predieri, "Plasma Chloroform Concentrations in Swimmers using Indoor Swimming Pools," *Arch. Environ. Health* **45**(3): 175-179 (1990).

10. K. T. Bogen, B. W. Colston and L. K. Machicao, "Dermal Absorption of Dilute Aqueous Chloroform, Trichloroethylene, and Tetrachloroethylene in Hairless Guinea Pigs," *Fund. Appl. Toxicol.* **18**, 30-39 (1992).

11. W. K. Jo, C. P. Weisel and P. J. Lioy, "Routes of Chloroform Exposure and Body Burden from Showering with Chlorinated Tap Water," *Risk Analysis* **10** (4), 575-580 (1990).

12. D. L. Morgan, S. W. Cooper, D. L. Carlock, J. J. Sykora, B. Sutton, D. R. Mattie and J. N. McDougal, "Dermal Absorption of Neat and Aqueous Volatile Organic Chemicals in the Fischer 344 Rat," *Environ. Res.* **55**: 51-63 (1991).

13. M. S. Islam, L. Zhao, J. N. McDougal and G. L. Flynn, "Uptake of Chloroform by Skin during Short Exposures to Contaminated Water," *Risk Analysis* (in press).

14. U.S. Environmental Protection Agency (EPA), "National Organic Monitoring Survey," Tech. Supp. Div. Off. of Water Supply, Cincinnati, Ohio (1977).

15. M. Gibaldi and D. Perrier, *Pharmacokinetics*, 2nd edition (Marcel Dekker, Inc., New York, 1982).

16. PCNONLIN, version 4.2, Scientific Consulting Inc., Apex, NC, USA.

17. W. L. Chiou, "Quantitation of Hepatic and Pulmonary First-Pass Effect and its Implications in Pharmacokinetic Study. I. Pharmacokinetics of Chloroform in Man," *J. Pharmacokin. Biopharm.* **3**(3):193-201 (1975).

18. B. J. Fry, T. Taylor and D. E. Hathway, "Pulmonary Elimination of Chloroform and its Metabolite in Man," *Arch. int. Pharmacodyn Ther.* **196**: 98-111 (1972).

19. J. R. Withey, B. T. Collins and P. G. Collins, "Effect of Vehicle on the Pharmacokinetics and Uptake of Four Halogenated Hydrocarbons from the Gastrointestinal Tract of the Rat," *J. Appl. Toxicol.* 3(5): 249-253 (1983).

20. J. R. Withey and B. T. Collins, "Chlorinated Aliphatic Hydrocarbons used in the Foods Industry: The Comparative Pharmacokinetics of Methylene Chloride, 1, 2 Dichloroethane, Chloroform and Trichloroethylene after I.V. Administratio in the Rat," *J.. Environ. Pathol. Toxicol.* 3: 313-332 (1980).

21. R. A. Corley, A. L. Mendrala, F. A. Smith, D. A. Staats, M. L. Gargas, R. B. Conolly, M. E. Andersen and R. H. Reitz, "Development of a Physiologically Based Pharmacokinetic Model for Chloroform," *Toxicol. Appl. Pharmacol.* 103: 512-527 (1990).

22. U.S. Environmental Protection Agency (EPA), "Dermal Exposure Assessment: Principles and Applications," Report No. EPA/600/8-91/011B, Office of Health and Environmental Assessment, U.S. EPA, Washington, D.C. 20460 (1992).

23. T. E. McKone and R. A. Howd, "Estimating Dermal Uptake of Nonionic Organic Chemicals from Water and Soil. Part 1. Unified Fugacity-based Models for Risk Assessments, *Risk Analysis* 12, 543-557 (1992).

24. R. L. Cleek and A. L. Bunge, "A New Method for Estimating Dermal Absorption from Chemical Exposure. 1. General Approach," *Pharm. Res* 10 (4), 497-506 (1993).

25. K. Bergman, "Whole-Body Autoradiography and Allied Tracer Techniques in Distribution and Elimination Studies of Some Organic Solvents," *Scand. J. Work Environ. & Health* 5(Suppl. 1): 1-263 (1979).

26. E. N. Cohen and N. Hood, "Application of Low-Temperature Autoradiography to Studies of the Uptake and Metabolism of Volatile Anesthetics in the Mouse. I. Chloroform," *Anesthesiology* 30(3): 306-314 (1969).

27. B. Löfberg and H. Tjälve, "Tracing Tissues with Chloroform-Metabolizing Capacity in Rats," *Toxicology* 39: 13-35 (1986).

Table I.   Pharmacokinetic parameters for the absorption and disposition of chloroform obtained by applying graphical methods (a) to the experimental data and the best estimates by fitting the data (b) to compartmental models using PCNONLIN.

IV Infusion (c) (3 - Compartment Disposition)

	α (min⁻¹)	β (min⁻¹)	K (min⁻¹)	Cmax (µg/ml)	Tmax (min)	AUC $_{0-\infty}$ (µg/ml/min)
Graphical Methods	-------	0.027	0.011	235.68	30	14843.37
Best-fit Estimate	0.16	0.019	0.005	238.10	30	--------

Dermal (2 - Compartment Disposition)

	Ka (min⁻¹)	α (min⁻¹)	K (min⁻¹)	Lag Time (min)	Cmax (µg/ml)	Tmax (min)	AUC $_{0-\infty}$ (µg/ml/min)
Graphical Methods	0.114	0.033	0.009	4.00	95.78	30.75	7973.91
Best-fit Estimate	0.050	0.035	0.008	7.59	87.83	32.98	--------

(a) Graphical Methods included the Method of Residuals, Trapezoidal Rule and visual inspection.

(b) Blood concentration data were weighted by a factor of 1/conc. before model-fitting.

(c) Infusion Rate was 0.633 mg/0.167 ml/min over 30 minutes.

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Table II. Comparison of the amount (mg) of chloroform dermally taken up by the hairless rat (over 464.4 cm^2) as estimated by using different equations (22-24) with our found values.

Exposure Time (min)	EPA (1992) (22)	McKone and Howd (23)	Cleek and Bunge(24)	Islam et al. (13) (total uptake found)	This Paper (systemic uptake found) *	Islam et al. (13) (only skin uptake) **
4	1.26	2.06	0.71	9.6		1.0
8	1.78	2.24	1.00	12.0		1.5
15	2.44	3.01	1.37	19.2		1.4
30	3.45	4.65	1.93	29.4	10.21	1.4

* Found by direct blood sampling.

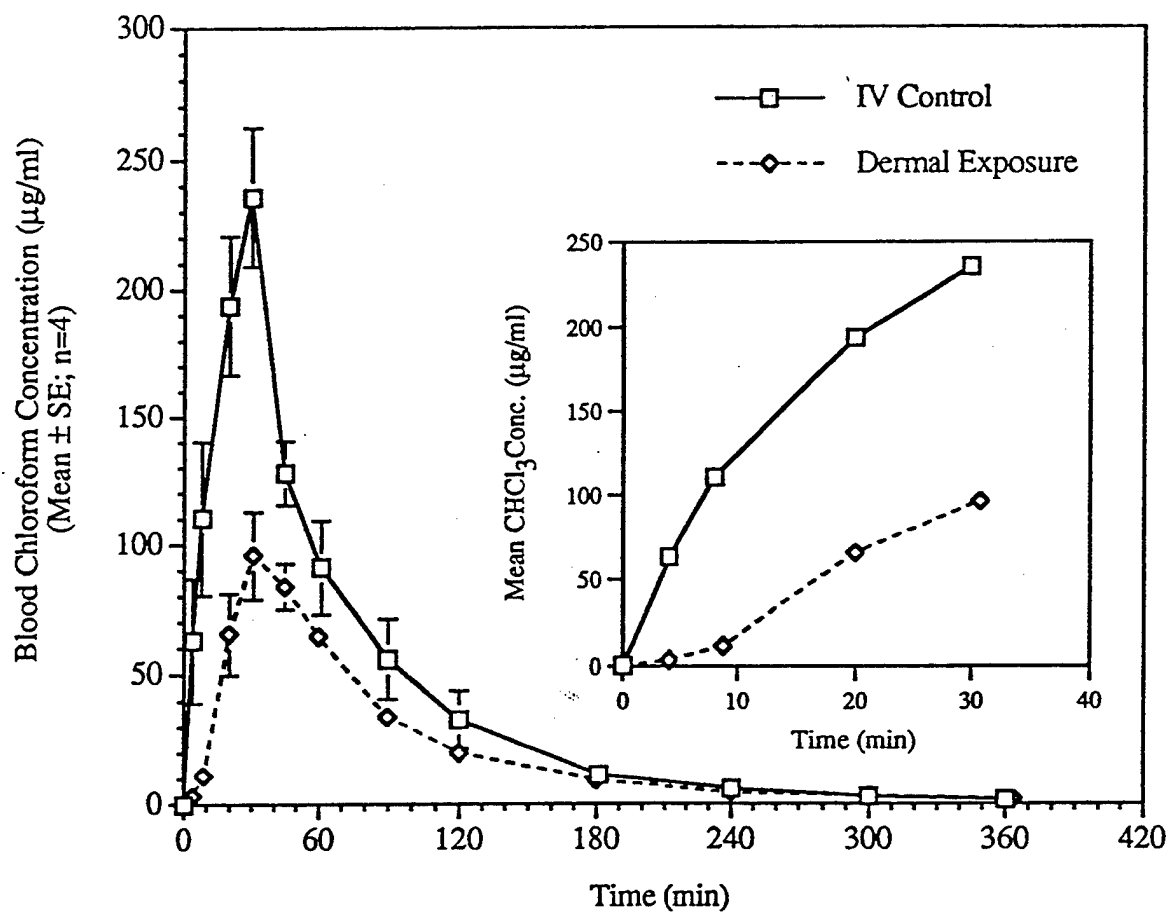** Amount recovered by tape-stripping only from the stratum corneum.

Fig. 1. Blood concentration-time profiles for chloroform after IV infusion of 19 mg (in 5 ml aqueous solution) over 30 min and after dermal exposure of the hairless rats to a 0.44 mg/ml aqueous solution for 30 min at 36°C. Each data point represents average ($\pm$ SE) of four experiments.
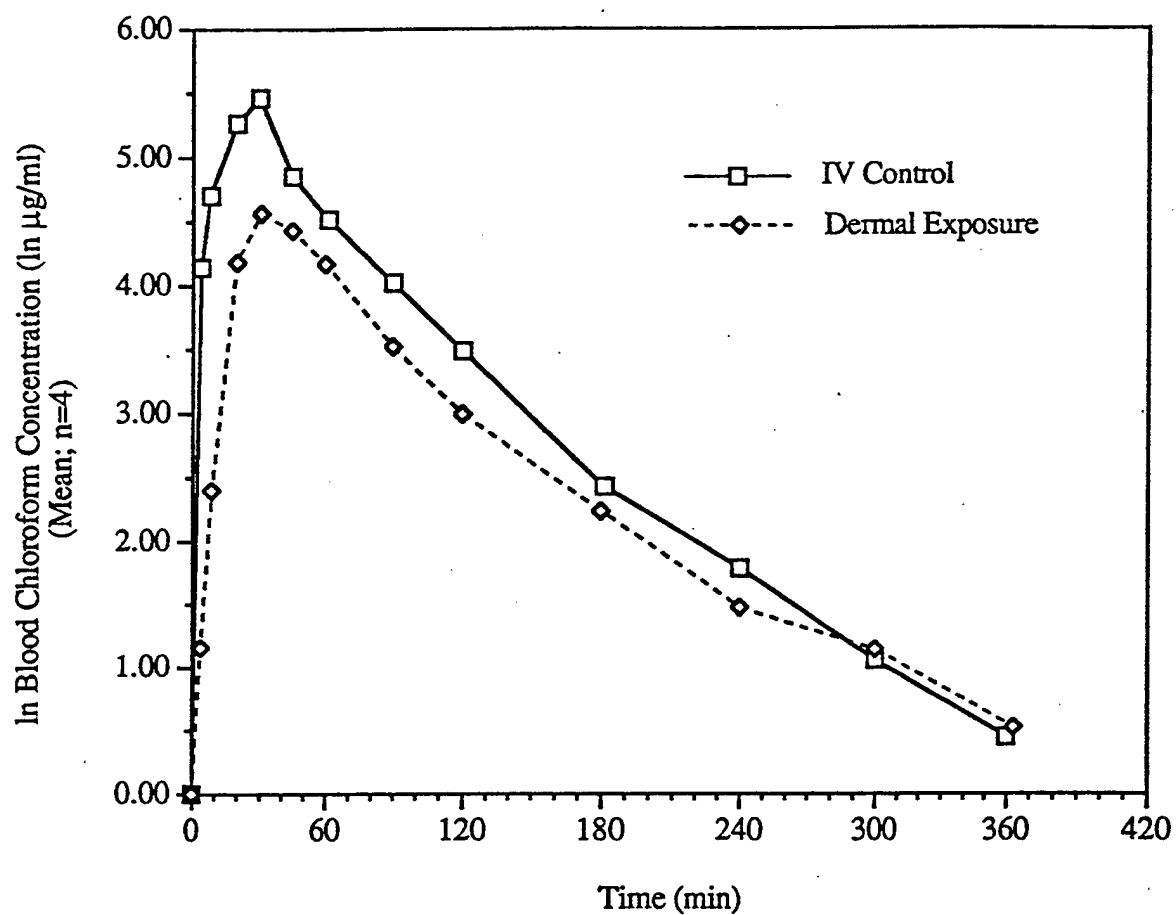
Fig. 2. Semilog plots of the same blood chloroform concentration vs. time data as used in Fig. 1 in order to delineate compartmental disposition characteristics.